Review

Anticancer Activity of Natural Bioactive Compounds against Human Carcinoma Cell Lines-A mini review

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Abstract

Cancer is one of the most stern health dilemmas engulfing 7 million lives every year globally. It is group of more than 100 diseases. Conventional cancer therapies burdened the disease crippled patient with lethal side effects and are also very expensive. Therefore, the demand to use alternative approaches in treatment of cancer is escalating. Plants of earth are massive reservoir of bioactive compounds. Plant derived compounds such as protoapigenone, casticin, moursin, gypenosides VN5, kalopanaxsaponin A, sapindoside B7, maclekarpine C are foremost anticancer agents, due to their unique structure and sophisticated mechanism of action. The present review is aimed at compiling data on promising classes of bioactive compounds from medicinal plants and their anti cancer mechanism of action against different human carcinoma cell lines.

Keywords

Alkaloid, Apigenin, Cell lines, Flavonoid, Moursin, Saponin.


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Introduction

Cancer is atrocious disease. It is second principal motive of mortality worldwide. Cancer is characterized by invasion, apoptotic death, angiogenesis, overexpression of cells, dysregulation of cell signaling pathways and metastasis (AGGARWAL & al [1]). Cancer cells may assault tissues which are in close proximity and may broaden through blood flow and lymphatic scheme to other parts of body (NAGARANI & al [2]). It causes more than six million deaths each year in the world (NISA & al [3]). Consistent with global cancer statistics issued by the American Cancer Society, daily 20,000 deaths were reported worldwide from cancer, in 2007. By 2050, 27 million new cancer cases and 17.5 million cancer deaths are predicted to crop up in the world [4].

Chief motives of cancer are viruses, environmental exposure (e.g. Ionizing and UV radiations), life style factors (e.g. high fat, low fiber diets, tobacco), medication (e.g. alkylating agents, immunosuppressant) and genetic factors, e.g. inherited mutations, cancer causing genes (MAMBET & al [5]). Moreover, When production of reactive oxygen species (e.g. superoxide anion radicals, hydroxyl radicals and hydrogen peroxide) is higher in body as compared to enzymatic and non-enzymatic antioxidants (e.g. superoxide dismutase, glutathione peroxidase, ascorbic acid, α-tocopherol, flavonoids, phenolics etc.) then this inequity accompanies cell harm and escort a vast range of degenerative diseases. Thus, reactive oxygen species are one of root of cancer.

Conventional modalities for treatment of cancer such as radiation therapy, chemotherapy, immunodilution and surgery has limited success as evident by high morbidity, toxicity, indiscriminate nature, intense site specificity, permanent disfigurement (NAWAB & al [6]). These reasons impart a strong momentum to researchers that there is an imperative need of new cancer management.

Plants are earliest associates of mankind. They have been employed for medical issues since 60,000 years ago. 250,000 species of medicinal plants persist on earth, out of which more than one thousand plant species are assessed to have significant anticancer activity (MUKHERJEE & al [7]), but there is still huge amount of work to be done on these lines. Affirmative effects of plants in anticancer therapy are attributed to profile of bioactive compounds in them.

Although bioactive components of plants include immense array of compounds which might impend or inhibit the inception of cancer (KOSANIC & al [8]). But flavonoids, e.g. apigenin, baicalin, genistein, catechin, saponins e.g. platycodon D, polyphenyllin D, Saikosaponin A and Tubeimoside I) and alkaloids e.g. camptothecin, vincristine, vinblastine, berbamine are promising anti-cancer agents (KOSANIC & al [8]; CHANG & al [9]).

Thus keeping in view the anticancer potential of promising classes of bioactive compounds, the present review compiles data on anticancer prospective of latest alkaloids, flavonoids and saponins extracted from terrestrial plants and their mechanism of action against various human carcinoma cell lines.

Flavonoids

Flavonoids (biflavonoids) polyphenolic plant compounds, own a common isoflavones, flavonols, flavanones and flavanons (REN & al [10]). They are present in flowers, roots, stems, fruits, vegetables, barks, nuts and herbal remedies (GALAT & al [11]). More than 4000 naturally occurring flavonoids have been reported (KUMAR & al [12]). They explore anticancer potential by apoptosis induction, anti-angiogenesis (REN & al [10]), topoisomerase inhibition (CONSTANTINOU & al [13]), accumulation of p53 (CIOBOTARU & al [14]), securing DNA from oxidative harm, triggering carcinogen detoxifying system, hindering carcinogen activation cell cycle arrest (LI & al [15]), improved expression of c-fos and c-myc (CHOI & al [16]) and inhibition of phosphatidyl-inositol 3-kinase (AGULO & al [17]). Thus it may be suggested that flavonoids are potential anticancer agents.

Four natural flavonoids casticin, artegenin, quercetin and 5, 30-dihydroxy-6,7,40-trimethoxy flavanone have been isolated from Vitex rotundifolia Linne fil. (KOYAKAWA & al [18]). Increase in number of cells as a result of growth and reproduction of similar cells by cell division is cause of cancer and is termed as cell proliferation. Anti proliferative activity of casticin in KB cell lines derived from human squamous cell carcinoma of the oral floor is excellent at low doses as compared to other flavonoids isolated from Vitex rotundifolia Linne fil. DNA histograms from flow cytometric analysis revealed that casticin noticeably arrest cell cycle at G2-M in KB cells.

Moreover, immunostaining explored that casticin exhibited marked antimitotic activity by disrupting mitotic spindles in KB cell lines. Anti-tumor activity of paclitaxel in KB cell lines is also reported (GARCIA & al [19]).

Many studies have suggested that flavonoids paly a defensive role in averting breast cancer (COORAY & al [20]). Three natural flavonoids epicatechin, proanthocyanidin B2 and proanthocyanidin B4 have been isolated from Litchi chinensis Sonn. (Litchi) pericarp (ZHAO & al [22]). Epicatechin, proanthocyanidin B2 and proanthocyanidin B4 are also previously reported from Litchi chinensis Sonn. (ZHAO & al [21]). Anti-breast cancer activities of Litchi chinensis Sonn. pericarp flavonoids have been evaluated in terms of proliferation of human breast cancer cell line MCF-7 by MTT assay (SUN & al [23]). Cytotoxic activities of epicatechin and proanthocyanidin B2 are higher in MCF-7 cells than proanthocyanidin B4. Authors also reported that paclitaxel, a reference anticancer agent, exhibited stronger cytotoxic activities than epicatechin and proanthocyanidin B2.

Protoapigenone, a natural flavonoid isolated from Thelypteris torresiana (Gaud) showed excellent cytotoxic activity in gynecological cancer cells especially MDAH-2774 and SKOV3 (ovarian cancer cells) approximated by XTT assay (CHANG & al [24]). Cytotoxic action of protoapigenone is credited to induction of apoptosis by
activating caspase-3 and cleaving the intact PARP. The cleavage of PARP is one of the salient feature of apoptosis and is activated by caspase-3, which is regulated by Bcl-2 family proteins (NICHOLSON & al [25]). Authors also employed immunohistochemical analysis in protoapigenone treated tumor specimens to notice elevated level of cell death is inappropriate (either too little or too much), then it is motive of various types of cancer. Failure of apoptosis is another cause of cancer. Cytotoxic effect of morsin against HeLa cells may be attributed to fact that morsin induced apoptosis by activating caspase-3,8 and 9 proteins and inhibiting nuclear factor kB signaling (LEE & al [26]). Induction of apoptosis is also reported by other flavonoids (CONSTANTINOU & al [27]).

Flavonoids of Teucrium ramosissimum inhibited proliferation of malignant cells at various levels. Apigenin, 4,7-dimethoxy apigenin, cirsimaritin and genkwanin have been reported in T. ramosissimum (SGHAIER & al [28]). Strongest activity on proliferation of human chronic myelogenous leukemia (K562) cells has been attained with apigenin followed by 4,7-dimethoxy apigenin, cirsimaritin, explored by MTT assay (POLYDORO & al [29]). Excellent cytotoxicity of apigenin is attributed to presence of hydroxyl group at C-4' and C-7', but presence of methoxy groups at same positions in 4,7-dimethoxy apigenin lessen its cytotoxicity. It is also evident from previous studies that position of hydroxyl group manipulate the confirmation of molecule and alter growth inhibitory consequences (KAWALI & al [30]). Apigenin is also recently tested on S2-013 and CD18 (human pancreatic cancer) cell lines and proves to be inhibitor of glucose transporter and phosphoinositide 3-kinase/Akt pathway (MELSTROM & al [31]). Very poor antiprolferative activity is scrutinized on human chronic myelogenous K562 cells by genkwanin.

**SAPONINS**

Saponis are surface active, non volatile, triterpene or steroid glycosides located in extensive array of plants and certain aquaculture (MAN & al [32]). They possess wide spectrum of biological and pharmacological properties such as anti diabetic (NORBERG & al [33]), anti atherosclerotic (ZHANG & al [34]), hemolytic, molluscidal (SPRAG & al [35]), anti inflammatory (LI & al [36]). Anticancer potential of saponins is attributed to inhibition of NF-Kb activation (HUANG & al [37]), induction of apoptosis in carcinoma cells (TROUILLAS & al [38]), cell cycle arrest (LIU & al [39]), inhibition of cell growth and DNA synthesis, mitotic arrest (KAUYEUNG & al [40]), antiproliferation, transcriptional activation of NSAID-activated gene (CHEN & al [41]) and others (KY & al [42]).

Seven dammarane-type saponins, gypenosides VN1-VN7, are reported from total saponin extract of Gynostemma pentaphyllium aerial parts (XING & al [43]). Isolated saponins exhibited moderate cytotoxic activity against HL-60 (leukemia), A549 (lung), HT-29 (colon), MCF-7 (breast), and SK-OV-3 (ovary) carcinoma cells. Cytotoxic effect is assessed by determining metabolic activity using MTT assay. Significant antitumor action of total saponin extract of Gynostemma pentaphyllum is also previously reported. Authors reported that only the IC$_{50}$ of gypenosides VN5 (IC$_{50}$ = 19.8 ±1.0) against MCF-7 cells is close to reference anticancer compound, mitoxantrone (IC$_{50}$ = 11.0 ± 0.8). Cytotoxic activity of gypenosides is accredited to induction of apoptosis in adenocarcinoma cells. Induction of apoptosis by gypenosides is also previously reported (WNAG & al [44]). Moreover, the mechanism of apoptosis induced by gypenosides in human colon cancer cells through the mitochondrial dependent pathways and activation of capase-3 is also on record (CHEN & al [45]). IC$_{50}$ values also explored that all the seven isolates (Gypenosides VN1-VN7) are weakly active or inactive against all the five carcinoma cell lines.

Three triterpene saponins, androsacin, ardisiacrispin A and saxifragifolin A have been isolated from Androsace integra (DONG & al [46]). Ardisiacrispin A and saxifragifolin A are also in literature records (WALTHO & al [47]). Cytotoxicity assay (WANG & al [48]) of three triterpene saponins against human hepatoma cell line (HepG2) illustrated that ardisiacrispin A has IC$_{50}$ of 1.56 μM Pro-apoptotic and microtubule disassembly effects of ardisiacrispin (A plus B) on human hepatoma Bel-7402 cells are previously reported (LYN & al [49]).

Three steroidal and two furostanol saponins, named as sarsaparilloside B, sarsaparilloside C, sarsaparilloside, parallin and D20(22)-sarsaparilloside have been detached from roots of Smilax sp. (CHALLINOR & al [50]). Antiprofleerative activity of five saponins against normal fibroblasts and different cancer cell lines have been evaluated. D20(22)-sarsaparilloside from roots of Smilax sp. showed highest cytotoxic effect against breast tumor (MCF-7), melanoma (MM96L) and leukemia (K562) cancer cell lines. Moreover, sarsaparilloside C, of Smilax sp. exhibited potent cytotoxic effect against colon tumor (HT29) cancer cell line. D20(22)-sarsaparillosid causes inhibition of cell proliferation by cell lysis as judged by appearance of greatly enlarged cytoplasm. Studies have demonstrated that saponins showed cytotoxicity of cancer cell lines by several mechanisms, such as mitotic inhibition, DNA replication, repair and growth factor signaling, apoptosis, cell lysis and others (CHEUNG & al [51]; TROUILLAS & al [52]; MAN & al [53]).

Three steroidal saponins named as penogenin-3-O-α-L-rhamnopyranosyl(1----2)-β-D-glucopyranoside, penogenin-3-O-α-L-rhamnopyranosyl(1----4)-α-L-rhamnopyranosyl (1----4)-[α-L-rhamnopyranosyl(1----2)]-β-D-glucopyranoside and 24-a-hydroxyl-penogenin-3-O-α-L-rhamnopyranosyl(1----2)-[α-L-arabinofuranosyl (1----4)]-β-D-glucopyranoside are isolated from Paris polyphylla Smith var. yunnanensis (MAN & al [54]). Antitumor activity of three saponins is elucidated by considering inhibition of growth and proliferation of HepG2 cells in dose dependant and time dependant manner.
and induction of apoptosis. Induction of apoptosis by steroidal saponin, polyphyllin D in drug resistant HepG2 cells is previously reported (CHEUNG & al [55]). Cell death pattern in HepG2 cells is addressed after treatment with three penogenin steroidal saponins individually, for 48 h at 20µM concentration followed by staining with aridine orange and ethidium bromide. Various morphological changes that occur in HepG2 cells are identified by fluorescence microscopy. In early stages of apoptosis cell shrinkage and pyknosis may be visualized by light microscopy. Pyknosis is result of chromatin condensation and is characteristic feature of apoptosis. Moreover, nuclear fragmentation, appearance of apoptotic bodies, high permeability of cell membrane and a window for both stained aridine orange and ethidium bromide is also visualized, which is characteristic of apoptosis.

Fourteen triterpene saponins including three new ones, 3β,6β,23-trihydroxyolean-12-oic acid 3-O-α-L-arabinopyranoside, kalopanaxsaponin L and kalopanaxsaponin M have been segregated from stem bark of Kalopanax pictus (QUANG & al [56]). Saponins from leaves of Kalopanax pictus are also in scientific reports. Cytotoxic activity of isolated saponins has been experienced against human cancer cell lines including leukemia (HL-60), colon (HCT-116), and breast (MCF-7), using MTT colorimetric method (SCUDIERO & al [57]). Most potent cytotoxic activity is exhibited by compound sapindoside B7, kalopanaxsaponin A and hederagenin 3-O-α-L-arabinopyranoside against HL-60, HCT-116, and MCF-7 carcinoma cell lines respectively. Cytotoxic activity of the oleanane-type triterpene saponins may be endorsed to free carboxyl group.

Five steroidal saponins have been segregated from Tribulus terrestris (SU & al [58]). Steroidal saponin from fruits of Tribulus terrestris are also previously reported (WANG & al [59]). Cytostatic activity of five isolated compounds against leukemia cancer cell lines (HL-60) is evaluated as IG50 values, which stands for inhibition of cell growth by 50%. IG50 values indicated that five saponins of Tribulus terrestris exhibited cytostatic attitude by suppressing cell growth and multiplication.

Two new oleanane triterpenoid saponins, xanthonhuskiside A and xanthonhuskiside B are detached from the husks of Xanthoceras sorbilofia (LI & al [60]). Growth inhibitory activities of xanthonhusksid A and xanthonhuskiside B against human leukemic monocyte lymphoma U937 cell line and human prostate cancer PC-3 cell line are estimated by trypan-blue and MTT methods. Cytotoxic acetylated triterpene saponin from the husks of Xanthoceras sorbilofia are already in literature collection (CHAN & al [61]). Xanthonhuskiside B illustrated moderate growth inhibitory activity against human leukemic monocyte lymphoma U937 cell line with IG50 value of 82.85±1.58µM. But xanthonhuskiside B against human prostate cancer PC-3 cell line and xanthonhuskiside A against both cell lines are inactive because their IG50 values are more than 100µM. Both the xanthonhuskiside A and B have glycosidation at C-28 which is pessimistic to anti tumor activity as previously suggested (LI & al [62]).

**Alkaloids**

Alkaloids (nitrogen-containing natural molecules independently of the basic character of the nitrogen) are copious secondary metabolites in plants and stand for one of the most widespread class of compounds endowed with multiple pharmacological properties (STEVIGNY & al [63]). They explored anticancer activity by inhibiting cell proliferation, inducing cell apoptosis (Programmed cell death characterized by discrete morphological processes) (HUANNG & al [64]), causing mitotic arrest during cell cycle, exhibiting antineoplastic activity by inhibition of DNA topoisomerase I and triggering mitochondrial pathway of apoptosis which escorts cancer cell death (CHIU & al [65]).

Five new dihydrobenzophenanthridine alkaloids along with ten known benzophenanthridine/dihydrobenzophenanthridine derivatives and a known amide are isolated from roots of Macleaya microcarpa (DENG & al [66]). New isolated alkaloids are named as maclekarpine A–E and their cytotoxicity is evaluated against five human carcinoma cell lines incorporating colon (HCT-8), liver (Bel-7402), stomach (BGC-823), ovarian (A2780) and lung (A549) by using MTT assay. IC50 values showed that excellent cytotoxicity of maclekarpine A and maclekarpine D is monitored against BGC-823 cell lines, while that of maclekarpine C and maclekarpine E is reported against A2780 and A549 carcinoma cell lines, respectively. Maclekarpine A is active only against human stomach cancer cell line (BGC-823). Maclekarpine B is found inactive against all the tested cell lines. Cytotoxic activity bioassay of known dihydrobenzophenanthridine alkaloids revealed that dihydrochelerythrine and dihydroosanguinarine (ZHANG & al [67]) methoxilated at C-6 displayed excellent results against HCT-8, A2780 and Bel-7402 cell lines.

Bioassay-guided fractionation of a CH2Cl2 extract from roots of Nauclea orientalis escorted the isolation of two new isomeric indole alkaloids, named as naucleorals A and naucleorals B (SUCHAEM & al [68]). Indole alkaloids from bark of Nauclea orientalis are also on record (ZHANG & al [69]). Dimeric indole alkaloids cause disruption of microtubules, dissolution of mitotic spindles and metaphase arrest in dividing cells. Cytotoxicity of naucleorals A and naucleorals B is assessed against KB (human epidermoid carcinoma) and HeLa (human cervical carcinoma) cell lines by colorimetric method (SICHAEM & al [70]). *In vitro* anticancer activity of indole alkaloids in ammonial extracts of Nauclea orientalis is also in literature (ERDELMEIER & al [71]). Excellent cytotoxic approach is examined by naucleorals A (IC50=4.0 µg/ml) against Hela cells, which is close to cytotoxic value of reference anticancer compound, adriamycin (IC50= 0.018 µg/ml).
Naucleorals B exhibited moderate cytotoxicity against HeLa and KB cells. Naucleorals A is inactive against KB cells.

Six voisinyl–ibogan type bisindole alkaloids containing four new ones ervachinines A–D and ten known monoterpenoid indole alkaloids are described from the whole plant of *Ervatamia chinensis* (GUO & al [72]). Monoterpenoid indole and bisindole alkaloids with antitumor potential are previously reported from genus *Ervatamia* (HIRASAWA & al [73]). Cytotoxic attitude of four new bisindole alkaloids ervachinines A–D is monitored against human myeloid leukemia (HL-60), hepatocellular carcinoma (SMMC-7721), lung cancer (A-549), breast cancer (MCF-7), and colon cancer (SW480 cells) cell lines by using MTT method. IC_{50} values calculated by Reed and Muech’s method explored that ervachinines A and ervachinines C exhibited better inhibitory activities than reference anticancer compound cisplatin, against all five cell lines. Cytotoxicity of ervachinines C is superior than that of ervachinines D against HL-60, SMMC-7721 and MCF-7 cells. Bisindole alkaloids interact with tubulin, dimeric cellular protein and cellular metabolic functions resulting in mitotic arrest and cell crenelation. Ervachinine E is recently reported from *Ervatamia chinensis* (GUO & al [74]) and modest cytotoxicity by it is explored against human myeloid leukemia (HL-60), hepatocellular carcinoma (SMMC-7721), lung cancer (A-549), breast cancer (MCF-7) and colon cancer (SW480 cells).

One aporphine alkaloid and nine other compounds are isolated from flowers of *Goniothalamus laoticus* (LEKPHROM & al [75]). Cytotoxicity of aporphine alkaloid, nordicentrine, is evaluated against human epidermoid carcinoma (KB), human breast cancer (BC1 and MCF-7) and human small cell lung cancer (NCI-H187) cell lines employing the colorimetric method. nordicentrine, exhibited excellent cytotoxicities against KB and NCI-H187 cells. This is in accordance with results of who showed excellent cytotoxic activities against twelve cancer cell lines. Moreover, cytotoxicities of nordicentrine against KB and NCI-H187 cells are close to cytotoxic values of doxorubicin and ellipticine, used as reference anticancer agents. Cytotoxic assay indicated that nordicentrine showed very strong activity against human epidermoid carcinoma and human small cell lung cancer cell lines. Good cytotoxic activity is observed by nordicentrine against MCF-7 cells, while activity of nordicentrine against human breast cancer cell lines is not determined.

### Table. Anticancer mechanism of action of bioactive compounds

<table>
<thead>
<tr>
<th>Compound</th>
<th>Mechanism of Action</th>
<th>Human Cancer Cell Line</th>
<th>Reference</th>
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<tbody>
<tr>
<td>Protoapigenone</td>
<td>Induction of apoptosis by activating caspase-3 and cleaving the intact PARP</td>
<td>MDAH-2774 SKOV3 HeLa C33A 468 T47D</td>
<td>[24]</td>
</tr>
<tr>
<td>Apigenin</td>
<td>Antiproliferative action</td>
<td>K562</td>
<td>[33]</td>
</tr>
<tr>
<td>Moursin</td>
<td>Induction of apoptosis by activating capase-3,8 and 9 proteins &amp; Inhibition of nuclear factor kBa signaling</td>
<td>HeLa MCF-7 Hep3B</td>
<td>[33]</td>
</tr>
<tr>
<td>D20(22)-sarsaparillosid</td>
<td>Inhibition of cell proliferation by cell lysis</td>
<td>MCF-7 MM96L K562</td>
<td>[54]</td>
</tr>
<tr>
<td>Gypenosides VN5</td>
<td>Induction of apoptosis</td>
<td>HL-60 MCF-7 HT-29 A-549 SKOV-3</td>
<td>[47]</td>
</tr>
<tr>
<td>Ervachinines A-D</td>
<td>Mitotic arrest Cell crenelation</td>
<td>MCF-7 HL-60 A-549 SMMC-7721 SW480</td>
<td>[76]</td>
</tr>
</tbody>
</table>

### Conclusion

Plants are pipeline of biologically active and miscellaneous anticancer agents. Although a range of plants is screened for anticancer potential, but lot of work is to be done on these lines. In this article, we reviewed some plant based active compound having potential against vast range of human carcinoma cell lines, which will be used in future for development of drugs exclusive of any adverse effects.
References


